



## Sequentially aerated membrane biofilm reactors for autotrophic nitrogen removal: microbial community composition and dynamics

**Pellicer i Nàcher, Carles; Franck, Stephanie ; Gülay, Arda; Rusalleda, M.; Terada, A.; Abu Al-Soud, W.; Asser Hansen, M.; Sørensen, S. J.; Smets, Barth F.**

*Published in:*  
Microbial Biotechnology

*Link to article, DOI:*  
[10.1111/1751-7915.12079](https://doi.org/10.1111/1751-7915.12079)

*Publication date:*  
2014

*Document Version*  
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

*Citation (APA):*  
Pellicer i Nàcher, C., Franck, S., Gülay, A., Rusalleda, M., Terada, A., Abu Al-Soud, W., Asser Hansen, M., Sørensen, S. J., & Smets, B. F. (2014). Sequentially aerated membrane biofilm reactors for autotrophic nitrogen removal: microbial community composition and dynamics. *Microbial Biotechnology*, 7(1), 32-43.  
<https://doi.org/10.1111/1751-7915.12079>

---

### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

## Sequentially aerated membrane biofilm reactors for autotrophic nitrogen removal: microbial community composition and dynamics

Carles Pellicer-Nàcher,<sup>1</sup> Stéphanie Franck,<sup>1</sup> Arda Gülay,<sup>1</sup> Maël Rusalleda,<sup>2</sup> Akihiko Terada,<sup>3</sup> Waleed Abu Al-Soud,<sup>4</sup> Martin Asser Hansen,<sup>4</sup> Søren J. Sørensen<sup>4</sup> and Barth F. Smets<sup>1\*</sup>

<sup>1</sup>Department of Environmental Engineering, Technical University of Denmark, Building 113, Miljøvej, 2800 Kgs Lyngby, Denmark.

<sup>2</sup>Laboratory of Chemical and Environmental Engineering (LEQUIA-UdG), Facultat de Ciències, Institute of the Environment, University of Girona, Campus Montilivi s/n, E-17071, Girona, Catalonia, Spain.

<sup>3</sup>Department of Chemical Engineering, Tokyo University of Agriculture & Technology, Naka-cho 2-24-16, Koganei, 184-8588 Tokyo, Japan.

<sup>4</sup>Department of Biology, Section for Microbiology, University of Copenhagen, Sølvgade 83H, 1307 Copenhagen K, Denmark.

### Summary

**Membrane-aerated biofilm reactors performing autotrophic nitrogen removal can be successfully applied to treat concentrated nitrogen streams. However, their process performance is seriously hampered by the growth of nitrite oxidizing bacteria (NOB). In this work we document how sequential aeration can bring the rapid and long-term suppression of NOB and the onset of the activity of anaerobic ammonium oxidizing bacteria (AnAOB). Real-time quantitative polymerase chain reaction analyses confirmed that such shift in performance was mirrored by a change in population densities, with a very drastic reduction of the NOB *Nitrospira* and *Nitrobacter* and a 10-fold increase in AnAOB numbers. The study of biofilm sections with relevant 16S rRNA fluorescent probes revealed strongly stratified biofilm structures fostering aerobic ammonium oxidizing bacteria (AOB)**

in biofilm areas close to the membrane surface (rich in oxygen) and AnAOB in regions neighbouring the liquid phase. Both communities were separated by a transition region potentially populated by denitrifying heterotrophic bacteria. AOB and AnAOB bacterial groups were more abundant and diverse than NOB, and dominated by the *r*-strategists *Nitrosomonas europaea* and *Ca. Brocadia anammoxidans*, respectively. Taken together, the present work presents tools to better engineer, monitor and control the microbial communities that support robust, sustainable and efficient nitrogen removal.

### Introduction

The discovery of anaerobic ammonium oxidizing bacteria (AnAOB, aka anammox bacteria) two decades ago has launched a new phase in wastewater biotechnology. Many reactor concepts have been developed to take advantage of this functional group for the treatment of nitrogen (N)-rich waste streams. AnAOB can grow symbiotically with aerobic ammonium oxidizing bacteria (AOB) in biofilms with redox gradients, allowing the conversion of equimolar mixtures of ammonium ( $\text{NH}_4^+$ ) and nitrite ( $\text{NO}_2^-$ ) to nitrogen gas ( $\text{N}_2$ ) without the addition of organic carbon (Terada *et al.*, 2011). In membrane-aerated biofilm reactors (MABRs) such redox gradient conditions can establish in a counter-diffusion mode, with oxygen ( $\text{O}_2$ ) entering the biofilm through the membrane–biofilm interface and  $\text{NH}_4^+$  diffusing from the liquid phase into the biofilm at its surface. We have recently shown that this biofilm reactor configuration can effectively support autotrophic N removal from synthetic waste streams at a lower energy, spatial, and environmental footprint than is feasible by conventional (i.e. based on co-diffusion) biofilm technologies (Pellicer-Nàcher *et al.*, 2010; Gilmore *et al.*, 2013).

$\text{NO}_2^-$  is a central intermediate in autotrophic N conversions: it is the product of AOB and is a necessary substrate for AnAOB. Therefore, metabolically active nitrite oxidizing bacteria (NOB) are undesirable during autotrophic N removal, as the oxidation of  $\text{NO}_2^-$  to nitrate ( $\text{NO}_3^-$ ), catalysed by NOB, would hamper AnAOB activity. Several strategies have been explored to suppress or control NOB growth in suspended growth or co-diffusion biofilm systems such as operation at elevated pH, low dissolved

Received 22 January, 2013; revised 21 June, 2013; accepted 26 July, 2013. \*For correspondence. E-mail bfm@env.dtu.dk; Tel. (+45) 45251600; Fax (+45) 45932850.  
doi:10.1111/1751-7915.12079

**Funding Information** Veolia Water and the Danish Agency for Science Technology and Innovation (FTP-ReSCoBiR) funded the present study. Maël Rusalleda was supported by the FI and BE (BE-2009-385) grant programmes from the Catalan Government (AGAUR).

oxygen (DO) concentration, or higher temperatures (Van Hulle *et al.*, 2010). However, none of these procedures has proven efficient to suppress NOB activity in MABRs (Wang *et al.*, 2009; Terada *et al.*, 2010). In large part, this difficulty in outcompeting NOB may be due to the localization of NOB, this microbial guild, in MABR biofilms. NOB, when present, grow in the aerobic regions of the biofilm (inner biofilm regions due to  $O_2$  diffusion across the biofilm base) where DO and  $NO_2^-$  concentrations are highest, and any changes applied to the liquid phase will have minimal effects. New approaches such as careful inoculum selection or implementation of cyclic aeration patterns have been explored and proven successful to moderate NOB activity in MABRs (Pellicer-Nàcher *et al.*, 2010; Terada *et al.*, 2010).

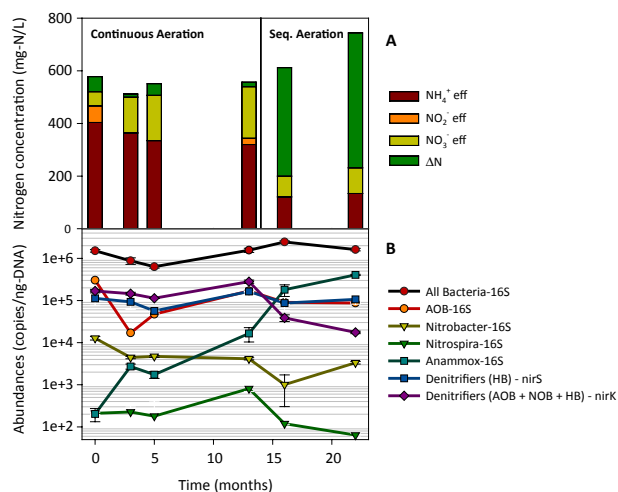
While the above studies have demonstrated the feasibility and identified suitable operational conditions for MABRs targeting autotrophic N removal (Pellicer-Nàcher *et al.*, 2010; Gilmore *et al.*, 2013), direct inspection of the microbial community structure and composition in the resulting MABR biofilms has been limited. Such inspection is not only necessary to assess the robustness of the process through the study of the diversity of the established microbial community, but also to deepen the understanding about how biofilms can be engineered for a certain operational purpose by applying selected operational strategies. In addition, information from direct biofilm inspection can be used to support or modify biofilm process models, where community compositions are easily predicted but rarely verified (Terada *et al.*, 2007; Lackner *et al.*, 2008).

Here, we present the first exhaustive characterization study of the structure and composition of microbial biofilms that support autotrophic N removal in MABRs (Pellicer-Nàcher *et al.*, 2010). We are especially keen to verify whether the imposed redox stratification results in the predicted ecological stratification of the involved functional groups, whether suppression of NOB and stimulation of AnAOB activity by sequential aeration is mirrored by the abundances of NOB and AnAOB, whether operational and reactor conditions have resulted in a more or less diverse set of functional guilds, and whether heterotrophic bacteria (HB) coexist in this autotroph-dominated community. Hence, we used a complementary set of molecular and microscopic tools to identify, quantify and assess the microbial diversity and structure of biofilms in a long-term operated MABR run under  $O_2$  limitation and treating a synthetic  $NH_4^+$  rich influent.

## Results and discussion

### Microbial dynamics during reactor operation

From inoculation to month 13, the reactor was operated in continuous aeration mode. The  $O_2$  to  $NH_4^+$  loading ratio



**Fig. 1.** Reactor performance and microbial community abundances during reactor operation. Month 0: reactor start-up after AnAOB inoculation. 23 months: reactor shutdown.

A. Averaged reactor performance during biomass sampling periods. Concentrations of  $NH_4^+$ ,  $NO_2^-$ ,  $NO_3^-$  and the N denitrified/assimilated  $\Delta N$  are stacked, yielding the  $NH_4^+$  concentration in the influent.

B. Population dynamics measured by real-time quantitative polymerase chain reaction (qPCR).

was controlled at the optimal value for complete  $NH_4^+$  conversion to  $N_2$  via the nitrification-anammox pathway (Terada *et al.*, 2007). Notwithstanding the imposed  $O_2$  limited operation, most of the  $NH_4^+$  was simply converted to  $NO_3^-$ , resulting in limited N removal (Fig. 1A). From month 13 onward,  $O_2$  was supplied in cycles of aeration/non-aeration, dramatically affecting reactor performance. Initially, strong  $NO_2^-$  accumulation was observed, followed by an increase in AnAOB activity and nearly maximal N removal (70%). In addition, at this point, minimal nitrous oxide ( $N_2O$ ) emissions were observed (Pellicer-Nàcher *et al.*, 2010).

Imposition of the sequential aeration regime caused clear shifts in the microbial community, as suggested by the results obtained from real-time quantitative polymerase chain reactions (qPCR) using relevant primers (Fig. 1B): the abundance of *Nitrobacter* spp. decreased by an order of magnitude. *Nitrobacter* spp. are considered *r*-strategist NOB (Schramm, 2003), whose presence is correlated with poor nitrification efficiencies (Terada *et al.*, 2010). Both 16S rRNA gene (Fig. 1B) and *nirA* (Fig. 1; in the Supporting Information, Fig. S1) targeted NOB quantifications were consistent. The gene copy numbers of *Nitrobacter* spp. increased by the end of the experiment (still under sequential aeration), but they did not negatively affect reactor performance (Fig. 1A). The abundance of the *Nitrospira* spp. was also severely affected by the onset of the sequential aeration: it decreased and dropped to the quantification limit until the end of the experiment. These data suggest that the

*Nitrospira* NOB, despite their lower abundance vis-à-vis *Nitrobacter* NOB, were responsible for the conversion of  $\text{NO}_2^-$  to  $\text{NO}_3^-$  during the antecedent continuous aeration phase. This NOB genus is known to thrive in environments with low  $\text{NO}_2^-$  concentrations (K-strategist, Schramm, 2003). The overall reduction in NOB abundance was mirrored by a significant increase in AnAOB numbers, as reflected by 16S rRNA gene (Fig. 1B) and *hzo*-targeted quantifications (Fig. S1). The exact reasons explaining the observed suppression of the NOB upon onset of the sequential aeration regime are unknown, but we hypothesize that AOB are less affected by the feast/famine conditions associated with cyclic aeration (Geets *et al.*, 2006) and display a higher affinity for  $\text{O}_2$  at low concentrations than NOB (Blackburne *et al.*, 2008), which would allow AOB to outcompete NOB for the transiently available limiting  $\text{O}_2$ . AOB are known to release hydroxylamine and NO during  $\text{O}_2$  transients, compounds which may have also inhibited NOB (Schmidt *et al.*, 2003; Noophan *et al.*, 2004; Kostera *et al.*, 2008). The increased  $\text{NO}_2^-$  availability (due to NOB suppression), and the postulated transiently available NO may also have enhanced  $\text{NO}_2^-$  uptake by AnAOB upon the onset of cyclic aeration (Jetten *et al.*, 2009; Kartal *et al.*, 2010). Although  $\text{NO}_2^-$  concentrations (in the bulk phase) attained values as high as  $200 \text{ mg-N l}^{-1}$ , they did not prevent the stimulation of AnAOB activity, in contrast with earlier studies that reported  $\text{NO}_2^-$  values as low as  $28 \text{ mg-N l}^{-1}$  to negatively affect their activity (van der Graaf *et al.*, 1996).

The density of denitrifying bacteria was approximated by quantifying the abundance of the *nirK* and *nirS* genes; *nirK* and *nirS* encode  $\text{NO}_2^-$  reductase enzymes in both heterotrophic denitrifying bacteria (*nirS* and *nirK*) or nitrifiers (*nirK*, Braker *et al.*, 1998; Casciotti and Ward, 2001; Heylen *et al.*, 2006). Onset of cyclic aeration caused a significant decrease in *nirK* abundance, while *nirS* remained relatively constant. Because AOB abundance remained fairly constant, this trend suggests a shift in the heterotrophic denitrifying guild, becoming more *nirS* abundant as AnAOB density and activity increased. The decrease in NOB, also known to contain *nirK* in their genome (Schreiber *et al.*, 2012), could have contributed as well to the observed drop. Prior work revealed that  $\text{N}_2\text{O}$  emissions from our and other MABRs were very low when AnAOB activity was high ( $\sim 0.015\%$  and  $< 0.001\%$   $\text{N-N}_2\text{O}/\text{N-load}$  during the aerated and non-aerated phase of an aeration cycle, respectively, Pellicer-Nàcher *et al.*, 2010), but can be very high when AnAOB activity is low ( $2\text{--}11\%$   $\text{N-N}_2\text{O}/\text{N-load}$ ; Gilmore *et al.*, 2013). Because  $\text{N}_2\text{O}$  emissions are, in part, caused by  $\text{NO}_2^-$  reductase (Zumft, 1997), it will be interesting to verify whether the dramatic drop in *nirK* abundance directly correlates with reduction in  $\text{N}_2\text{O}$  emissions.

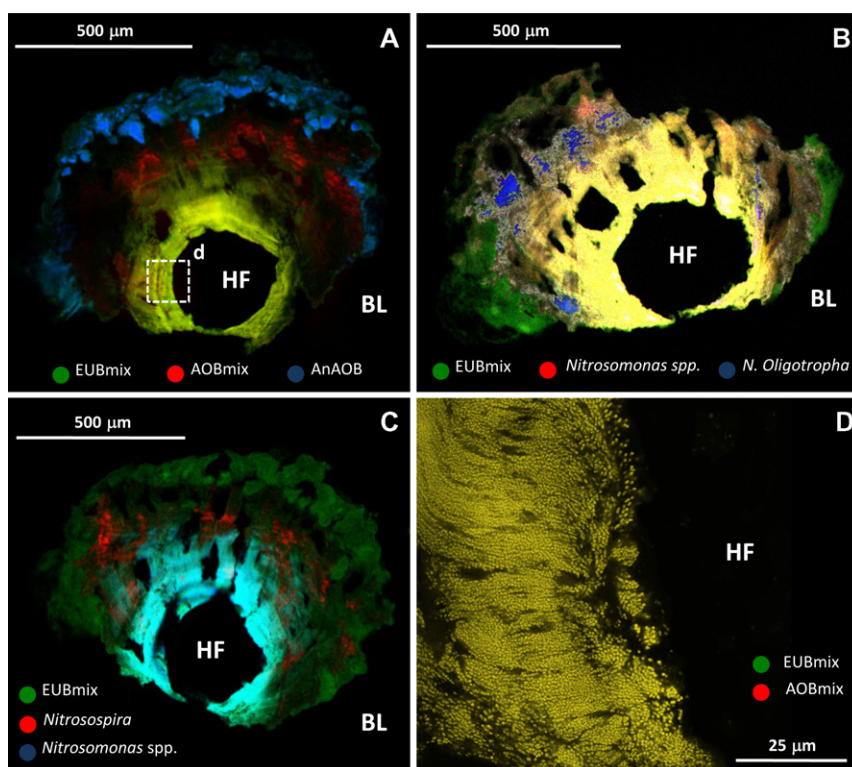
### Microbial community composition and architecture

After 23 months of operation (630 days), the developed biofilm exhibited a very pronounced radial stratification of the microbial community (Fig. 2A). The biofilm region adjacent to the hollow fibre membrane was clearly dominated by AOB, as indicated by the simultaneous signal from EUB – all bacteria – and AOB targeting probes (Table 1, combination 1). The thickness of the observed AOB layer ranged from 110 to 170  $\mu\text{m}$  across all analysed samples, comparable to the  $\text{O}_2$  penetration measured with microsensors during reactor operation here and in other nitrifying MABRs (Fig. S2, Terada *et al.*, 2010). Moreover, since the DO concentration in the bulk liquid was close to the detection limit of the DO probe used (Pellicer-Nàcher *et al.*, 2010), it could be concluded that the AOB group consumed most of the  $\text{O}_2$  transferred from the membrane, and mediated the partial conversion of the  $\text{NH}_4^+$  diffusing from the bulk liquid to  $\text{NO}_2^-$ . High magnification micrographs in this area revealed rod-shaped cells in very compact strata around the hollow fibre membranes (Fig. 2D). The average biofilm porosities calculated here were  $0.36 \pm 0.07$ , half of what is considered normal in co-diffusion biofilm systems (Zhang and Bishop, 1994). This packed and ordered structure of AOB is very different from the cauliflower-shaped clusters observed in other biofilm systems (Okabe and Kamagata, 2010), which may be caused from the pressure of upper biofilm strata onto the biofilm base, high growth velocities or the high competition for  $\text{O}_2$  and space in this region. Isolated cauliflower-shaped clusters could only be seen in biofilm regions distant from the membrane where DO concentrations were lower.

The use of probes with higher phylogenetic resolution (Table 1, combinations 2 and 3, Fig. 2B and C), indicated that halophilic and halotolerant *Nitrosomonas* spp. were the dominant AOB in the system. Halophilic and halotolerant *Nitrosomonas* spp. are known to outcompete other AOB species in environments with high substrate availability (Okabe and Kamagata, 2010). The high  $\text{NH}_4^+$  concentrations expected across the whole biofilm thickness (bulk concentrations ranged from 100 to  $400 \text{ mg-N l}^{-1}$ ) may hamper the ability of *Nitrospira* spp. to thrive even in biofilm areas with lower DO concentrations, as previously reported (Schramm *et al.*, 2000). Our observation is in agreement with the work by Terada *et al.* (2010), who noted that MABR biofilms populated by *Nitrosomonas* spp. rather than *Nitrospira* spp. supported higher  $\text{NO}_2^-$  production. Halophilic and halotolerant *Nitrosomonas* spp. are typically the most abundant AOB in co-diffusion biofilms performing autotrophic N removal (Sliekers *et al.*, 2003; Vázquez-Padín *et al.*, 2010; Liu *et al.*, 2012).

Microcolonies of the *N. oligotropha* lineage (Table 1, probe combination 3, Fig. 1B) were occasionally observed and randomly distributed within aerobic biofilm regions.





**Fig. 2.** Biofilm cross-sections hybridized with fluorescent probes targeting AOB and AnAOB. HF and BL stand for hollow fiber and bulk liquid respectively.

A. Biofilm cross-section hybridized with probes targeting All Bacteria (EUBmix), AOB and AnAOB.

B. Biofilm section hybridized with probes targeting All Bacteria (EUBmix), halophilic and halotolerant *Nitrosomonas* spp. and *N. oligotropha*.

C. Biofilm section hybridized with probes targeting All Bacteria (EUBmix), halophilic and halotolerant *Nitrosomonas* spp. and *Nitrosospira* spp.

D. Detail of Fig. 2A at the membrane–biofilm interface (d).

Although *N. oligotropha* is expected in environments of low substrate availability, *N. oligotropha* and *N. europaea* can coexist in aerobic regions of nitrifying biofilms when operated under dynamic aeration conditions (Gieseke *et al.*, 2001). In another MABR biofilm performing autotrophic N removal under continuous aeration *N. oligotropha* and halophilic and halotolerant *Nitrosomonas* spp. were also identified as the most abundant AOB (Gilmore *et al.*, 2013).

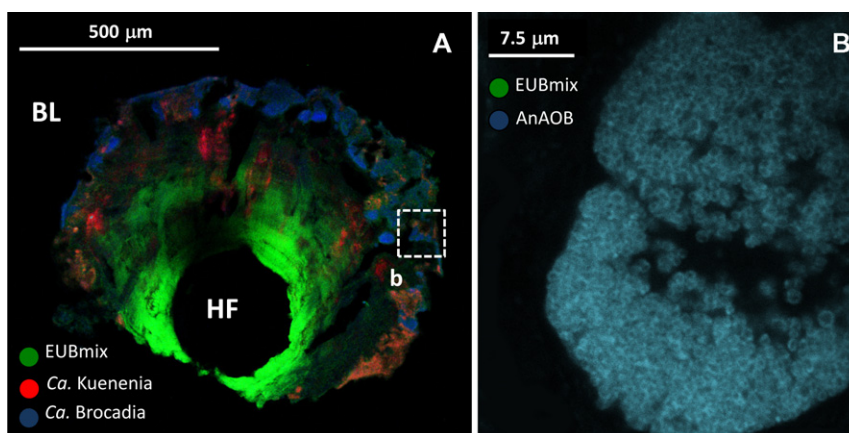
In that study however, *N. oligotropha* signals were observed in anaerobic strata parallel to the membrane surface, suggesting cell maintenance without growth, as *N. oligotropha* is, among other AOB, able to maintain its ribosome content even under famine conditions (Gieseke *et al.*, 2001).

AnAOB-positive signals were exclusively found close to the biofilm top, at least 300 μm away from the membrane

**Table 1.** Probe combinations used in the study (after Loy *et al.*, 2007; Okabe and Kamagata, 2010). Probe sequences and hybridization conditions for each probe are available in Table S1.

No.	Experimental Purpose	Fluorophore		
		FLUO (green)	Cy3 (red)	Cy5 (blue)
1.	Verify spatial distribution of AOB and AnAOB	EUBmix	Nso190-Nmo218-Cluster 6a192 <sup>a</sup>	Amx820
2.	Examine abundance of <i>Nitrosospira</i> spp. vs halophilic and halotolerant <i>Nitrosomonas</i> spp. (AOB)	EUBmix	Nsv443	NEU <sup>a</sup>
3.	Examine abundance of <i>N. oligotropha</i> vs halophilic and halotolerant <i>Nitrosomonas</i> spp. (AOB)	EUBmix	NEU <sup>a</sup>	Nmo218-Cluster 6a192 <sup>a</sup>
4.	Examine abundance of <i>Candidatus</i> Kuenenia spp. vs <i>Candidatus</i> Brocadia spp. (AnAOB)	EUBmix	Kst157	Ban162
5.	Examine abundance of <i>Nitrobacter</i> spp. vs <i>Nitrospira</i> spp. (NOB)	EUBmix	NIT3 <sup>a</sup>	Ntspa662 <sup>a</sup>

a. Probe requires a competitor



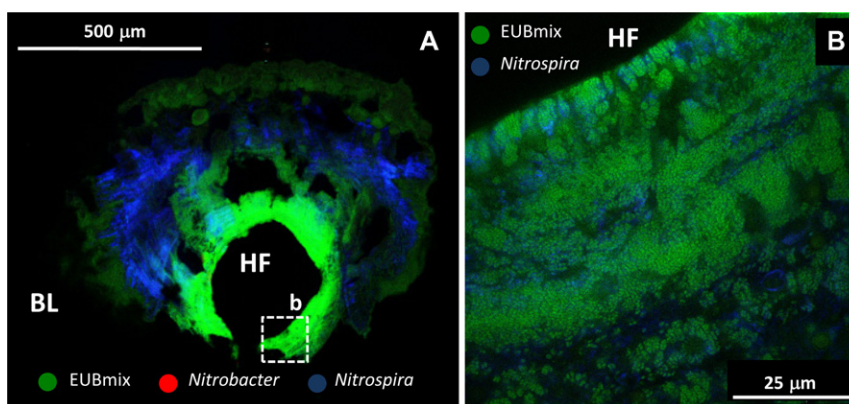
**Fig. 3.** Biofilm cross-sections hybridized with fluorescent probes targeting AnAOB. HF and BL stand for hollow fiber and bulk liquid respectively. A. Biofilm cross-section hybridized with probes targeting All Bacteria (EUBmix), *Ca. Brocadia*, and *Ca. Kuenenia*. B. Detail of Fig. 3-A in regions neighbouring the bulk liquid (b).

surface (Fig. 2A). Microsensor measurements confirmed that DO was almost absent in this region (Fig. S2), while high  $\text{NH}_4^+$  and  $\text{NO}_2^-$  concentrations were expected, given their concentrations in the bulk liquid (Fig. 1A). AnAOB microcolonies were spearhead- or oval-shaped with round edges. High magnification micrographs (Fig. 3B) revealed doughnut-shaped cellular morphology, characteristic of AnAOB (rRNA distributes around the central anammoxosome organelle, Kuenen, 2008).

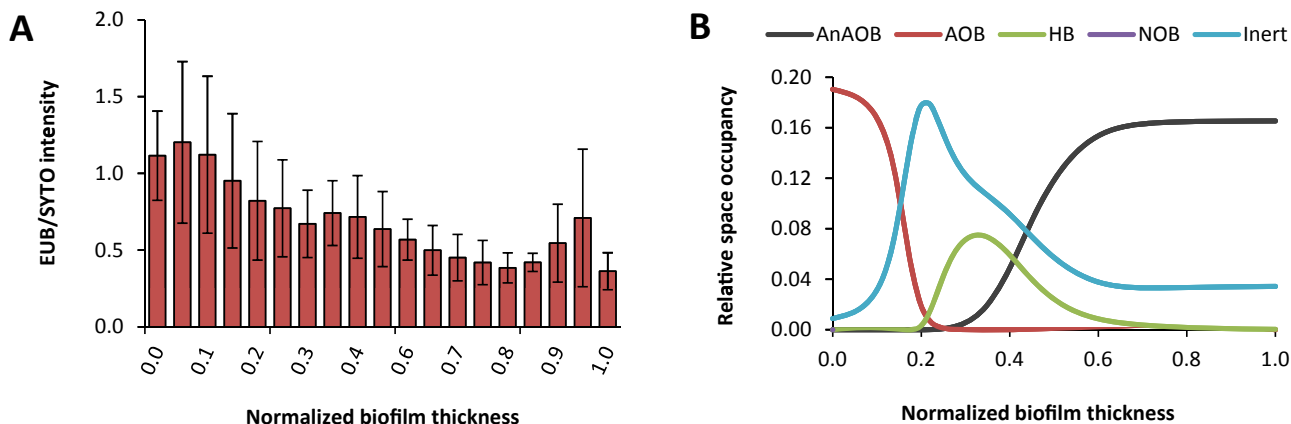
Although it is rare to find multiple AnAOB species in bioreactors performing autotrophic N removal (Hu *et al.*, 2010; Park *et al.*, 2010a), both *Ca. Brocadia* anammoxidans (dark cyan) and *Ca. Kuenenia stuttgartiensis* (light orange) were observed here. However, *Brocadia* signals were clearly more abundant (Fig. 3A, Table 1, combination 4). These two AnAOB have different postulated growth preferences: *Ca. Brocadia* seems to thrive in environments with high substrate availability (*r*-strategist), while *Ca. Kuenenia* finds its niche in environ-

ments with nutrient scarcity (*K*-strategist) (van der Star *et al.*, 2008). Microcolonies of both AnAOB lineages do not appear to be stratified within our biofilms. This observation might be due to the changing  $\text{NH}_4^+$  and  $\text{NO}_2^-$  concentrations during sequential aeration. In a continuously aerated MABR performing autotrophic N removal exclusively the *Ca. Brocadia* AnAOB lineage was observed (Gilmore *et al.*, 2013).

Overall reactor mass balance calculations suggested that, although substantial N removal was observed ( $5.5 \text{ g-N m}^{-2} \text{ day}^{-1}$ ), approximately 30% of the  $\text{NO}_3^-$  production was due to residual NOB activity ( $0.3 \text{ g-N m}^{-2} \text{ day}^{-1}$ ), while the remainder had been synthesized by AnAOB ( $0.7 \text{ g-N m}^{-2} \text{ day}^{-1}$ , Pellicer-Nàcher *et al.*, 2010). Here, NOB could still be detected in the biofilm (Table 1, combination 5, Fig. 4), even though they were clearly outnumbered by AOB in the aerobic biofilm regions. NOB in these biofilms were exposed to dual competitive pressure caused by fast growing AOB (consuming most  $\text{O}_2$ ) and AnAOBs (consum-



**Fig. 4.** Biofilm cross-sections hybridized with fluorescent probes targeting NOB. HF and BL stand for hollow fiber and bulk liquid respectively. A. Biofilm section hybridized with probes targeting All Bacteria (EUBmix), *Nitrobacter* and *Nitrospira*. B. Detail of Fig. 4A in a region close to the aeration membrane (b).



**Fig. 5.** Observed and predicted occurrence of metabolically active biofilm regions. Normalized biofilm thickness = 1 (biofilm top). Normalized biofilm thickness = 0 (membrane–biofilm interface).

A. Averaged relative activity profile with biofilm depth ( $n = 4$ ). Standard deviations represented with error bars in each column.

B. Relative space occupancy profile for the considered bacterial communities calculated by mathematical modelling in a previous study describing heterotrophic activity in MABR biofilms for autotrophic N removal (Lackner *et al.*, 2008).

ing the generated  $\text{NO}_2^-$ ; Terada *et al.*, 2010; Gilmore *et al.*, 2013). Only *Nitrospira*, a *K*-strategist NOB, was observed in the biofilm after fluorescent in-situ hybridization (FISH) inspection (Fig. 4A), while *Nitrobacter*, detectable via qPCR analyses, was not observed, suggesting their lower activity. In well-functioning co-diffusion autotrophic N removal biofilms, both NOB types have been found (Sliekers *et al.*, 2002; Park *et al.*, 2010a).

While *Nitrospira* signals appeared most abundant distant from the biofilm base, scattered *Nitrospira*-positive cells were also detected in the aerobic regions close to the biofilm–membrane interface (Fig. 4B). This observation is in contrast with a previous MABR study, which suggested that DO concentrations at the membrane–biofilm interface above  $63 \mu\text{M}$  result in a shift in the NOB community from *Nitrospira* to *Nitrobacter*, leading to a reduction in nitrification efficiencies (Downing and Nerenberg, 2008). In our system, DO concentrations at the membrane–biofilm interface were as high as  $300 \mu\text{M}$  (Fig. S2), and yet no *Nitrobacter* signals were observed. High nitrification turnover is therefore possible in MABRs under intensive  $\text{O}_2$  loading conditions if a cyclic aeration pattern is imposed.

Between the aerobic (AOB-signal rich) and anaerobic (AnAOB-signal rich) biofilm regions, a transitional zone existed where no conclusive phylogenetic assignments could be made based on the employed FISH probes. A non-sense probe (non-EUB) was successfully applied at the first stages of the study to rule out any autofluorescence or unspecific probe binding in this region. Additional analyses of metabolic activity were made by normalizing the cellular ribosomal content (using EUBmix signal as proxy) to the total cellular genomic content (using SYTO 60 signal as proxy) across the biofilm. This analysis yielded three tentative peaks (Fig. 5A); the peaks at the biofilm base (0) and top (1)

were indicative of the described AOB and AnAOB regions. A third additional peak in the centre of the biofilm (normalized biofilm thickness of 0.4) might indicate the presence of metabolically active cells in this transition region (Fig. 5A), even though the increase in signal intensity observed was not statistically significant, as could be concluded from the ANOVA test performed. Application of double-labelled oligonucleotide probes (DOPE probes, Stoecker *et al.*, 2010) did not improve the assignments (results not shown). Earlier modelling efforts in MABRs for autotrophic N removal predicted the accumulation of HB and large amounts of bacterial debris in this transition zone (Fig. 5B, Lackner *et al.*, 2008). Together with the qPCR results (showing abundant *nirS* numbers, indicative of heterotrophic denitrifiers), this suggests an anoxic transition zone populated by a heterotrophic denitrifying community. In addition, the non-specific and low probe signals observed suggest that these regions contained biomass debris and non-viable cells, consistent with the described model predictions.

#### Microbial community abundances

Quantitative image analysis of the presented FISH results revealed that AOB and AnAOB accounted for  $53 \pm 13\%$  and  $38 \pm 6\%$  of the biofilm, with  $9 \pm 8\%$  of the EUB signals not accounted for by either AOB or AnAOB (Table 1, combination 1). A separate analysis indicated a highly variable NOB fraction constituting  $11 \pm 10\%$  of all detected EUB signals (Table 1, combination 5). These AOB, NOB and AnAOB fractions are consistent with those observed by FISH analysis in co-diffusion biofilms performing autotrophic N removal (Liu *et al.*, 2012).

These qFISH results compare with those obtained by qPCR, for which AOB, NOB and AnAOB fractions were



**Table 2.** Relative abundance of functional microbial guilds after 630 days of operation assessed via different molecular techniques.

	Pyrosequencing <sup>a</sup>	qPCR	FISH
AOB	2.70 ± 0.87%	5.4 ± 0.2%	53.7 ± 13.2%
NOB	0.57 ± 0.12%	0.2 ± 0.2%	11.4 ± 10%
AnOB	60.00 ± 3.42%	25.0 ± 12.1%	38.2 ± 5.8

a. Taxonomy index of each functional guild was given in supplementary document (Table S1).

estimated at about 5, 0.2 and 25% of the total community population (considering that each cell contained a single copy of the genes targeted by qPCR primers, Table 2). Differences in sampling procedure may have impacted considerably the AOB and NOB results obtained by qPCR. While the entire biofilm is probed and imaged during FISH analysis (both biofilm and membrane were cryosectioned), only the biomass that could be scraped off the membranes was quantified by qPCR. As nitrifiers were preferentially present adjacent to the aeration membrane, AOB and NOB underestimation by qPCR was likely. Additionally, DNA extraction methods are known to display different yields in different types of bacterial species, which may bias substantially later qPCR analyses (Juretschko *et al.*, 2002; Foesel *et al.*, 2008). The targeted functional genes may be present in multiple copies per cell in the different biocatalysts studied, which could also lead to wrong estimates of the community fractions by qPCR calculated here (Pei *et al.*, 2010). Finally, small differences could also be explained by the fact that, oligonucleotide probes (for FISH) and primers (for qPCR) do not necessarily detect exactly the same phylogenetic clades (Hallin *et al.*, 2005).

The presented results, however, seem to converge in the fact that AOB and AnAOB are the most abundant microbial guilds. Model calibration and process operation can greatly benefit from the dual detection approach presented here. While FISH and microelectrode results would give information on the position and relative abundances of the functional microbial communities (Schramm *et al.*, 2000), qPCR performed on representative biomass samples would allow for routine observation measurements on the microbial dynamics of the system. These observations could alert about unwanted changes in the microbial community supporting the process and suggest the need of imposing process control to correct deviations from the desired set point (Park *et al.*, 2010a).

#### Diversity of community fractions assessed by deep sequencing

Microbial diversity of relevant functional guilds (AOB, NOB and AnAOB) was revealed by pyrosequencing the V3-V4 region of amplified community 16S rDNA. After implementation of quality control measures and denoising, 19 962

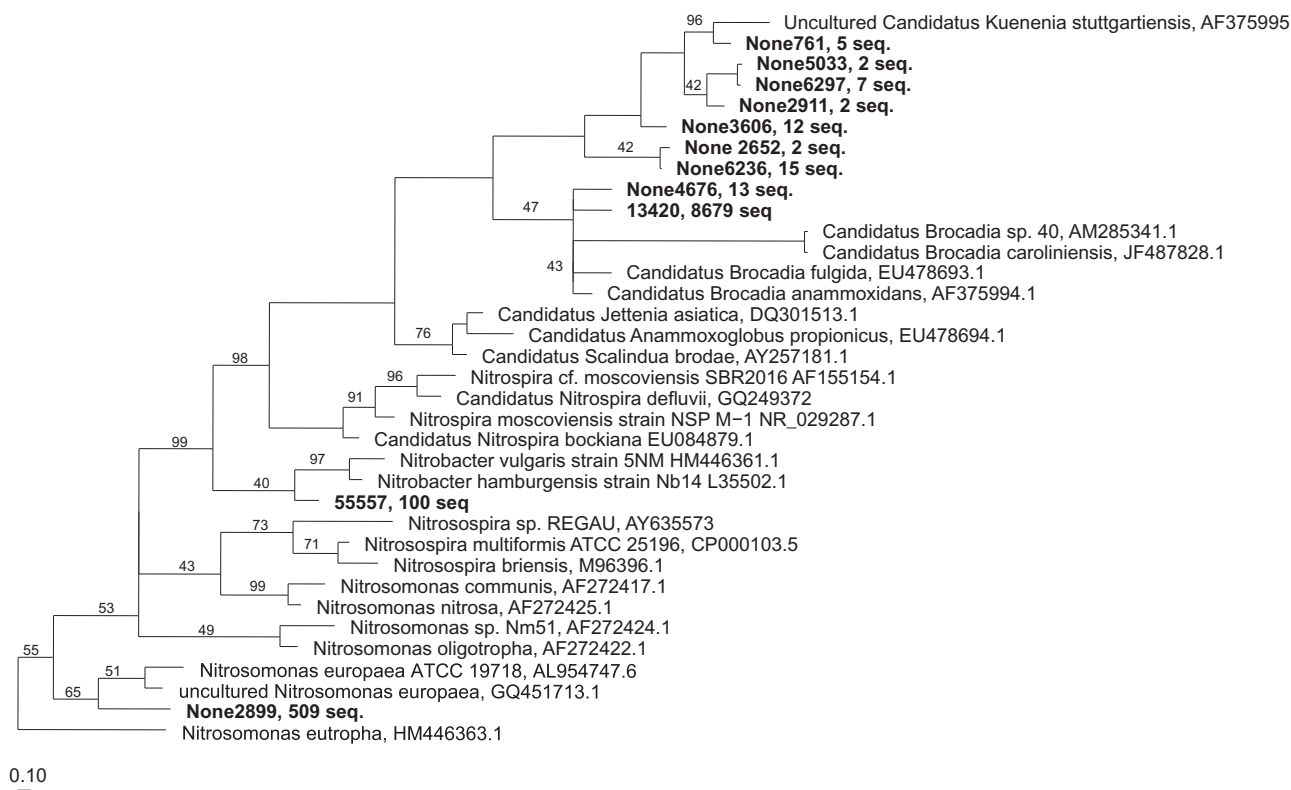
sequences were obtained from triplicate samples, which could be binned in 439 operational taxonomic units based on 97% phylogenetic similarity (OTU<sub>0.03</sub>). A total of 15, 2 and 6 OTUs<sub>0.03</sub> could be assigned to AnAOB, NOB and AOB respectively (Fig. 6). All three sequence libraries shared 117 OTUs<sub>0.03</sub> but were distinct due to abundant unshared genotypes (Fig. S3).

A remarkable diversity was detected within the AnAOB guild containing 15 OTUs<sub>0.03</sub>, while sampling depth may not yet have revealed the complete diversity. Sequences of both the *Kuenenia* and *Brocadia* lineages were detected. While species richness in both lineages was the same (8 each), *Brocadia* sequences were significantly more abundant (290:1), consistent with FISH observations. Sequences affiliated to the aerobic ammonium oxidizer *Nitrosomonas europaea* (509 seq.) and the NO<sub>2</sub><sup>-</sup> oxidizer *Nitrobacter hamburgensis* (100 seq.) were the dominant nitrifying bacteria in the studied MABR biofilms (Fig. 6). While the diversity of AOB was relatively high, species different to *N. europaea* represented only minor fractions of the AOB guild (most having a single sequence for each OTU<sub>0.03</sub>, occurring in only one of the replicates, Fig. S4). Higher microbial diversity is normally reported in systems with dynamic operation conditions (Rowan *et al.*, 2003).

Heterotrophic OTUs<sub>0.03</sub> (41) were found in all samples despite the absence of organic carbon in the synthetic feed. The most abundant HB sequences were mainly assigned to the *Xanthomonadaceae* (γ-Proteobacteria, 550 seq.) and *Clostridiaceae* (Firmicutes, 356 seq.) families. Our results are in agreement with the findings of a study applying microautoradiography combined with FISH (MAR-FISH) to unravel the patterns in the cross-feeding of microbial products originating from nitrifier decay to HB (Okabe *et al.*, 2005). In that study *Chloroflexi* and *Cytophaga-Flavobacterium* cluster cells were responsible for the degradation of slow biodegradable material (e.g. cells). Furthermore, members of the α- and γ-Proteobacteria (e.g. *Xanthomonadaceae*, typical in low carbon environments) metabolized other low-molecular weight organic substrates. Members of the *Clostridiaceae* family were not detected by their approach (gram positive bacterial require special fixation protocols), but, like *Xanthomonadaceae*, they are also known to degrade complex organic substrates and use NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> as electron acceptor in anaerobic environments (Finkmann *et al.*, 2000; Wüst *et al.*, 2011). CARD FISH may further assist in the identification of the organisms present in this intermediate region.

The fractions of AOB, NOB and AnAOB were identified based on the number of sequences detected by pyrosequencing. The results were further compared to those obtained by qPCR and FISH (Table 2). The abundance results obtained by pyrosequencing confirm the





**Fig. 6.** Phylogenetic tree of identified AOB, NOB and AnAOB sequences, and reference strains. Sequences from 16S rRNA libraries were assembled in OTUs based on 97% similarity. Distance matrices were computed with Jukes-Cantor method, and phylogenetic trees were rendered based on neighbour joining with bootstrap replication. Numbers at the branch nodes indicate bootstrap values over 40%. Number of identified sequences in each OTU is listed. OTUs with singular sequences are not shown. Full tree is available in Figure S4.

reduced NOB abundance with respect to AOB and AnAOB. The quantification of AOB and *Nitrospira* spp. was also affected with respect to qFISH. As previously indicated, such a divergence in results can arise by the difficulty in detaching the aerobic biofilm regions from the membrane or by a lower DNA extraction efficiency of the proposed treatment for these microbial species. The differences observed among PCR-based techniques could be related to the different amplification efficiency of universal primers (used in pyrosequencing) compared with family- and genus-specific primers (used in qPCR). Indeed, universal primers enhance the detection of highly abundant taxa but disfavour species represented by lower fractions (Gonzalez *et al.*, 2012).

## Conclusion

Imposition of sequential aeration was successful in the rapid suppression of NOB activity and stimulating and maintaining AnAOB activity in the MABR, at very high effective  $O_2$  loadings ( $4\text{--}14\text{g-}O_2\text{ m}^{-2}\text{ day}^{-1}$ ). qPCR-based analysis revealed the following most remarkable shifts in the biofilm composition: a strong and moderate decrease

in, respectively, the *nirK*, *Nitrospira* and *Nitrobacter* 16S rRNA gene abundance, and a strong increase in the AnAOB 16S rRNA gene abundance.

FISH analysis of the biofilms, after long-term sequential aeration, confirmed radial microbial stratification, with AOB at very high cellular densities in the  $O_2$ -rich areas close to the membrane surface, and AnAOB located close to the bulk liquid, separated by a transition region potentially harbouring denitrifying HB supported by decay products. Extant diversity assessed by using phylogenetically defined FISH probes revealed that halophilic and halotolerant *Nitrosomonas* spp., *Ca. Brocadia anammoxidans* and *Nitrospira* spp. were the most abundant lineages within the AOB, NOB and AnAOB guilds respectively. AOB and AnAOB constituted the most abundant community fractions, outnumbering NOB. Deep sequencing of the mature biofilm showed that the AOB guild was dominated by a single *N. europaea* OTU and confirmed a diverse AnAOB guild containing one most abundant *Brocadia* spp. OTU. Overall, our multipronged analysis confirmed that sequential aeration regimes can be used to successfully to engineer a microbial community to attain high-rate autotrophic N removal.

## Experimental procedures

### Samples

Samples were obtained from a MABR performing stable N removal at  $5.5 \text{ g-N m}^{-2} \text{ day}^{-1}$  via the nitrification-anammox pathway. This reactor housed 10 membrane bundles, each containing 128 30 cm long hollow-fibres (Model MHF3504, polyethylene/polyurethane, Mitsubishi Rayon, Tokyo, Japan). The reactor was inoculated in two steps with biomass from nitrifying (day -30) and AnAOB enrichment cultures (day 0), and was fed a synthetic  $\text{NH}_4^+$ -rich influent ( $100\text{--}500 \text{ mg-N l}^{-1}$ ), while air was passed through the fibre lumens ( $2.5\text{--}40 \text{ kPa}$  and  $2.5\text{--}60 \text{ l min}^{-1}$ ). After 390 days of operation the reactor was aerated in sequential cycles comprising aerated and non-aerated periods (Pellicer-Nàcher *et al.*, 2010). On days 90, 150, 390 and 480, biofilm samples were detached and removed from the reactor by inserting a Pasteur pipette through the installed sampling ports and aspirating biomass from several fibre bundles at several heights. In these samples, biofilm structure could not be preserved. More comprehensive and structurally intact biofilm samples were obtained sacrificially at the end of the reactor run (day 630).

### FISH analysis

Biofilm samples were collected after 630 days of operation by cutting one fibre at three different locations along its length (lower, middle and upper). Specimens containing both biofilm and membrane were fixed in 4% paraformaldehyde, embedded in OCT compound (Sakura Finetek Europe, Zoeterwoude, the Netherlands), frozen at  $-21^\circ\text{C}$ , cut in  $20 \mu\text{m}$ -thick sections by using a microtome, and mounted on gelatine-coated slides. Sectioned samples were dehydrated and sequentially probed with Fluorescein (FLUO), Cy3- and Cy5- tagged 16S rRNA probes (Sigma Aldrich, St Louis, MO, USA, Table 1), following procedures described elsewhere (Terada *et al.*, 2010). Biofilm structure was further studied by staining the prepared sections with a general fluorescent nucleic acid stain following manufacturer's specifications (SYTO 60, Invitrogen, Carlsbad, CA, USA).

Hybridized and stained sections were inspected with a confocal laser-scanning microscope (CLSM, Leica TCS SP5, Leica Microsystems, Wetzlar, Germany) equipped with an Ar laser (488 nm), and two HeNe lasers (543 and 633 nm). Gain and pinhole settings were tuned for each registered image in order to allow unsaturated exposure values for all detected pixels. At least five micrographs obtained from specimens hybridized with probe combinations 1 and 5 (Table 1) were used to quantify the abundance of AOB, AnAOB and NOB using the image analysis software Daime (Daims *et al.*, 2006). Images of SYTO stained biofilms were analysed with Leica AS AF Lite (Leica) and ImagePro (MediaCybernetics, Des Moines, IA, USA) software to determine biofilm porosities and probe intensity profiles along biofilm depth.

### DNA extraction

DNA was extracted from 0.5 g of biofilm mass (dry weight) using the MP FastDNA™ SPIN Kit (MP Biomedicals, Solon, OH, USA) according to manufacturer's instructions. The

quality of the extracted genomic DNA was checked by measuring its 260/280 nm absorbance ratio with a NanoDrop (ThermoFisher Scientific, Waltham, MA, USA) and was later stored at  $-20^\circ\text{C}$  in Tris-EDTA buffer until further processing.

### High-resolution tag-based 16S rRNA sequence library

Biofilm DNA sampled on day 630 was amplified with Phusion (Pfu) DNA Polymerase (ThermoFisher Scientific) and the 16S rDNA universal primers PRK341F (5'-C CTAYGGG RBGCAACAG-3') and PRK806R (5'-GG ACTACNNGGG TATCTAAT-3') (Yu *et al.*, 2005) with 25 annealing and elongation cycles. Detailed description of the PCR amplification conditions have been described before (Masoud *et al.*, 2011; Sundberg *et al.*, 2013). Amplified DNA was purified using the QIAquick PCR purification kit (Qiagen, Vento, the Netherlands) according to the manufacturer's protocol. In a second 15-cycle PCR round, barcodes and tags were added. All 16S rDNA fragments comprising the V3 and V4 hypervariable regions were pyrosequenced using a 454 FLX Titanium sequencer (Roche, Penzberg, Germany). Detailed description of the pyrosequencing preparation process can be found elsewhere (Farnelid *et al.*, 2011).

### Bioinformatic analyses

All raw 16S rDNA amplicons were processed and classified using the QIIME (<http://qiime.org/index.html>) software package (Caporaso *et al.*, 2010b). Chimera checking and denoising were performed with the software Ampliconnoise (Quince *et al.* 2011). Retrieved sequences were clustered at 97% evolutionary similarity and aligned against the Greengenes reference set (DeSantis *et al.*, 2006) using the Pynast algorithm (Caporaso *et al.*, 2010a). Taxonomy index of each functional guild is given in Table S2. Phylogenetic trees were created in ARB using library sequences of interest together with selected representatives from a small subunit (SSU) reference library (SSU Ref. Nr. 111 Silva). Statistical calculations and phylogenetic comparisons were carried out using R software (R Development Core Team, 2012).

### Quantitative PCR (qPCR)

Biofilm DNA sampled and extracted throughout the whole operational period (days 0, 90, 150, 390, 480 and 630) was subject to qPCR to determine the numbers of AOB, NOB, AnAOB and denitrifying bacteria, based on appropriate 16S rRNA targets or functional genes (Table 3). Details on the procedure can be found elsewhere (Pellicer-Nàcher *et al.*, 2010; Terada *et al.*, 2010).

### Microelectrode measurements

Oxygen Clark-type microsensors were constructed to measure DO concentrations within the biofilm. Preparation, calibration and microsensor measurement were performed as previously described (Revsbech, 1989; Pellicer-Nàcher *et al.*, 2010).

### Acknowledgements

The authors would like to thank Ms Lene Kirsten Jensen and Dr Marlene Mark Jensen for their assistance in con-

**Table 3.** Primers and conditions used for the quantification of bacterial numbers by qPCR.

Primers	Target Organism	Target molecule	Sequence (5'-3')	Reference
1055f 1392r	All Bacteria	Eubacterial 16S rRNA gene	ATG GCT GTC GTC AGC T ACG GGC GGT GTG TAC	Lane, 1991; Ferris <i>et al.</i> , 1996
CTO189fa/b CTO189fc RT1r	β-proteobacterial AOBs	16S RNA gene	GGA GRA AAG CAG GGG ATC G GGA GGA AAG TAG GGG ATC G CGT CCT CTC AGA CCA RCT ACT G	Kowalchuk <i>et al.</i> , 1997; Hermansson and Lindgren, 2001
FGPS872f 1269r	<i>Nitrobacter</i> NOB	16S RNA gene	CTA AAA CTC AAA GGA ATT GA TTT TTT GAG ATT TGC TAG	Degrange and Bardin, 1995
Nspra675f 746r	<i>Nitrospira</i> NOB	16S RNA gene	GCG GTG AAA TGC GTA GAK ATC G TCA GCG TCA GRW AYG TTC CAG AG	Graham <i>et al.</i> , 2007
F1370f1 F2843r2	<i>Nitrobacter</i> / <i>Nitrococcus</i> NOB	nrxA gene	CAG ACC GAC GTG TGC GAA AG TCC ACA AGG AAC GGA AGG TC	Poly <i>et al.</i> , 2008; Wertz <i>et al.</i> , 2008
Amx809f Amx1066r	AnAOB	16S RNA gene	GCC GTA AAC GAT GGG CAC T ATG GGC ACT MRG TAG AGG GGT TT	Tsushima <i>et al.</i> , 2007
hzoqf hzoqr	AnAOB	Hzo gene	CAT GGT CAA TTG AAA GRC CAC C GCC ATC GAC ATA CCC ATA CTS	Park <i>et al.</i> , 2010b
cd3aF R3cd	Denitrifying bacteria	nirS gene	AAC GYS AAG GAR ACS GG GAS TTC GGR TGS GTC TTS AYG AA	Throback <i>et al.</i> , 2004
F1aCu R3Cu	Denitrifying bacteria	nirK gene	ATC ATG GT(C/G) CTG CCG CG GCC TCG ATC AG(A/G) TTG TGG TT	Hallin and Lindgren, 1999

ducting the presented qPCR analyses, Dr. Frank Schreiber for executing the presented microsensor measurements, Ms Gizem Multlu for her assistance in the FISH work and Dr Arnaud Dechesne for his valuable comments on the manuscript. Ms Annie Ravn Petersen is greatly thanked for her valuable help with the cryostat microtome.

### Conflict of Interest

None declared.

### References

- Blackburne, R., Yuan, Z.G., and Keller, J. (2008) Partial nitrification to nitrite using low dissolved oxygen concentration as the main selection factor. *Biodegradation* **19**: 303–312.
- Braker, G., Fesefeldt, A., and Witzel, K.-P. (1998) Development of PCR primer systems for amplification of nitrite reductase genes (*nirK* and *nirS*) to detect denitrifying bacteria in environmental samples. *Appl Environ Microbiol* **64**: 3769–3775.
- Caporaso, J.G., Bittinger, K., Bushman, F.D., DeSantis, T.Z., Andersen, G.L., and Knight, R. (2010a) PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinformatics* **26**: 266–267.
- Caporaso, J.G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F.D., Costello, E.K., *et al.* (2010b) QIIME allows analysis of high-throughput community sequencing data. *Nat Methods* **7**: 335–336.
- Casciotti, K., and Ward, B. (2001) Dissimilatory nitrite reductase genes from autotrophic ammonia-oxidizing bacteria. *Appl Environ Microbiol* **67**: 2213–2221.
- Daims, H., Lucker, S., and Wagner, M. (2006) Daime, a novel image analysis program for microbial ecology and biofilm research. *Environ Microbiol* **8**: 200–213.
- Degrange, V., and Bardin, R. (1995) Detection and counting of *Nitrobacter* populations in soil by PCR. *Appl Environ Microbiol* **61**: 2093–2098.
- DeSantis, T.Z., Hugenholtz, P., Larsen, N., Rojas, M., Brodie, E.L., Keller, K., *et al.* (2006) Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol* **72**: 5069–5072.
- Downing, L.S., and Nerenberg, R. (2008) Effect of oxygen gradients on the activity and microbial community structure of a nitrifying, membrane-aerated biofilm. *Biotechnol Bioeng* **101**: 1193–1204.
- Farnelid, H., Andersson, A.F., Bertilsson, S., Al-Soud, W.A., Hansen, L.H., Sørensen, S., *et al.* (2011) Nitrogenase gene amplicons from global marine surface waters are dominated by genes of non-cyanobacteria. *PLoS ONE* **6**: e19223.
- Ferris, M.J., Muyzer, G., and Ward, D.M. (1996) Denaturing gradient gel electrophoresis profiles of 16S rRNA-defined populations inhabiting a hot spring microbial mat community. *Appl Environ Microbiol* **62**: 340–346.
- Finkmann, W., Altendorf, K., Stackebrandt, E., and Lipski, A. (2000) Characterization of N<sub>2</sub>O-producing *Xanthomonas*-like isolates from biofilters as *Stenotrophomonas nitritireducens* sp. nov., *Luteimonas mephitis* gen. nov., sp. nov. and *Pseudoxanthomonas broegbermensis* gen. nov., sp. nov. *Int J Syst Evol Microbiol* **50**: 273–282.
- Foesel, B.U., Gieseke, A., Schwermer, C., Stief, P., Koch, L., Cytryn, E., *et al.* (2008) *Nitrosomonas* Nm143-like ammonia oxidizers and *Nitrospira marina*-like nitrite oxidizers dominate the nitrifier community in a marine aquaculture biofilm. *FEMS Microbiol Ecol* **63**: 192–204.
- Geets, J., Boon, N., and Verstraete, W. (2006) Strategies of aerobic ammonia-oxidizing bacteria for coping with nutrient and oxygen fluctuations. *FEMS Microbiol Ecol* **58**: 1–13.
- Gieseke, A., Purkhold, U., Wagner, M., Amann, R., and Schramm, A. (2001) Community structure and activity dynamics of nitrifying bacteria in a phosphate-removing biofilm. *Appl Environ Microbiol* **67**: 1351–1362.
- Gilmore, K.R., Terada, A., Smets, B.F., Love, N.G., and Garland, J.L. (2013) Autotrophic nitrogen removal in a membrane-aerated biofilm reactor under continuous aeration: a demonstration. *Environ Eng Sci* **30**: 38–45.

- Gonzalez, J.M., Portillo, M.C., Belda-Ferre, P., and Mira, A. (2012) Amplification by PCR artificially reduces the proportion of the rare biosphere in microbial communities. *PLoS ONE* **7**: e29973.
- van der Graaf, A.A.V., De Bruijn, P., Robertson, L.A., Jetten, M.S.M., Kuenen, J.G., van De Graaf, A.A., *et al.* (1996) Autotrophic growth of anaerobic ammonium-oxidizing micro-organisms in a fluidized bed reactor. *Microbiology* **142**: 2187–2196.
- Graham D.W., Knapp C.W., Van Vleck E.S., Bloor K., Lane T.B., and Graham C.E. (2007) Experimental demonstration of chaotic instability in biological nitrification. *ISME J* **1**: 385–393.
- Hallin, S., and Lindgren, P.-E. (1999) PCR Detection of genes encoding nitrite reductase in denitrifying bacteria PCR detection of genes encoding nitrite reductase in denitrifying bacteria. *Appl Environ Microbiol* **65**: 1652–1657.
- Hallin, S., Lydmark, P., Kokalj, S., Hermansson, M., Sörensson, F., Jarvis, Å., *et al.* (2005) Community survey of ammonia-oxidizing bacteria in full-scale activated sludge processes with different solids retention time. *J Appl Microbiol* **99**: 629–640.
- Hermansson, A., and Lindgren, P.-E. (2001) Quantification of ammonia-oxidizing bacteria in arable soil by real-time PCR. *Appl Environ Microbiol* **67**: 972–976.
- Heylen, K., Gevers, D., Vanparys, B., Wittebolle, L., Geets, J., Boon, N., *et al.* (2006) The incidence of *nirS* and *nirK* and their genetic heterogeneity in cultivated denitrifiers. *Environ Microbiol* **8**: 2012–2021.
- Hu, B., Zheng, P., Tang, C., Chen, J., van der Biezen, E., Zhang, L., *et al.* (2010) Identification and quantification of anammox bacteria in eight nitrogen removal reactors. *Water Res* **44**: 5014–5020.
- van Hulle, S.W.H., Vandeweyer, H.J.P., Meesschaert, B.D., Vanrolleghem, P.A., Dejans, P., and Dumoulin, A. (2010) Engineering aspects and practical application of autotrophic nitrogen removal from nitrogen rich streams. *Chem Eng J* **162**: 1–20.
- Jetten, M.S.M., van Niftrik, L., Strous, M., Kartal, B., Keltjens, J.T., Op den Camp, H.J.M., *et al.* (2009) Biochemistry and molecular biology of anammox bacteria. *Crit Rev Biochem Mol Biol* **44**: 65–84.
- Juretschko, S., Loy, A., Lehner, A., and Wagner, M. (2002) The microbial community composition of a nitrifying-denitrifying activated sludge from an industrial sewage treatment plant analyzed by the full-cycle rRNA approach. *Syst Appl Microbiol* **25**: 84–99.
- Kartal, B., Tan, N.C.G., van de Biezen, E., Kampschreur, M.J., van Loosdrecht, M.C.M., and Jetten, M.S.M. (2010) Effect of nitric oxide on anammox bacteria. *Appl Environ Microbiol* **76**: 6304–6306.
- Kostera, J., Youngblut, M.D., Slosarczyk, J.M., and Pacheco, A.A. (2008) Kinetic and product distribution analysis of NO center dot reductase activity in *Nitrosomonas europaea* hydroxylamine oxidoreductase. *J Biol Inorg Chem* **13**: 1073–1083.
- Kowalchuk, G.A., Stephen, J.R., De Boer, W., Prosser, J.I., Embley, T.M., and Woldendorp, J.W. (1997) Analysis of ammonia-oxidizing bacteria of the beta subdivision of the class *Proteobacteria* in coastal sand dunes by denaturing gradient gel electrophoresis and sequencing of PCR-amplified 16S ribosomal DNA fragments. *Appl Environ Microbiol* **63**: 1489–1497.
- Kuenen, J.G. (2008) Anammox bacteria: from discovery to application. *Nat Rev Microbiol* **6**: 320–326.
- Lackner, S., Terada, A., and Smets, B.F. (2008) Heterotrophic activity compromises autotrophic nitrogen removal in membrane-aerated biofilms: results of a modeling study. *Water Res* **42**: 1102–1112.
- Lane, D.J. (1991) 16S/23S rRNA sequencing. In *Nucleic Acid Techniques in Bacterial Systematics*. Stackebrandt, E., and Goodfellow, M. (eds). New York, USA: Wiley, pp. 115–175.
- Liu, T., Li, D., Zeng, H., Li, X., Liang, Y., Chang, X., *et al.* (2012) Distribution and genetic diversity of functional microorganisms in different CANON reactors. *Bioresour Technol* **123**: 574–580.
- Loy, A., Maixner, F., Wagner, M., and Horn, M. (2007) probeBase – an online resource for rRNA-targeted oligonucleotide probes: new features 2007. *Nucleic Acids Res* **35**: D800–D804.
- Masoud, W., Takamiya, M., Vogensen, F.K., Lillevang, S., Al-Soud, W.A., Sørensen, S.J., *et al.* (2011) Characterization of bacterial populations in Danish raw milk cheeses made with different starter cultures by denaturing gradient gel electrophoresis and pyrosequencing. *Int Dairy J* **21**: 142–148.
- Noophan, P., Figueroa, L.A., and Munakata-Marr, J. (2004) Nitrite oxidation inhibition by hydroxylamine: experimental and model evaluation. *Water Sci Technol* **50**: 295–304.
- Okabe, S., and Kamagata, Y. (2010) Wastewater treatment. In *Environmental Molecular Microbiology*. Liu, W.T., and Jansson, J.K. (eds). Norfolk, UK: Caister Academic Press, pp. 191–210.
- Okabe, S., Kindaichi, T., and Ito, T. (2005) Fate of <sup>14</sup>C-labeled microbial products derived from nitrifying bacteria in autotrophic nitrifying biofilms. *Appl Environ Microbiol* **71**: 3987–3994.
- Park, H., Rosenthal, A., Jezek, R., Ramalingam, K., Fillos, J., and Chandran, K. (2010a) Impact of inocula and growth mode on the molecular microbial ecology of anaerobic ammonia oxidation (anammox) bioreactor communities. *Water Res* **44**: 5005–5013.
- Park, H., Rosenthal, A., Ramalingam, K., Fillos, J., and Chandran, K. (2010b) Linking community profiles, gene expression and N-removal in anammox bioreactors treating municipal anaerobic digestion reject water. *Environ Sci Technol* **44**: 6110–6116.
- Pei, A.Y., Oberdorf, W.E., Nossa, C.W., Agarwal, A., Chokshi, P., Gerz, E., *et al.* (2010) Diversity of 16S rRNA genes within individual prokaryotic genomes. *Appl Environ Microbiol* **76**: 3886–3897.
- Pellicer-Nàcher, C., Sun, S.P., Lackner, S., Terada, A., Schreiber, F., Zhou, Q., *et al.* (2010) Sequential aeration of membrane-aerated biofilm reactors for high-rate autotrophic nitrogen removal: experimental demonstration. *Environ Sci Technol* **44**: 7628–7634.
- Poly, F., Wertz, S., Brothier, E., and Degrange, V. (2008) First exploration of *Nitrobacter* diversity in soils by a PCR cloning-sequencing approach targeting functional gene *nxrA*. *FEMS Microbiol Ecol* **63**: 132–140.



- Quince, C., Lanzen, A., Davenport, R.J., and Turnbaugh, P.J. (2011) Removing noise from pyrosequenced amplicons. *BMC Bioinformatics* **12**: 1–18.
- R Development Core Team (2012) R: A language and environment for statistical computing. R Foundation for Statistical computing, Vienna, Austria.
- Revsbech, N.P. (1989) An oxygen microsensor with a guard cathode. *Limnol Oceanogr* **34**: 474–478.
- Rowan, A.K., Moser, G., Gray, N., Snape, J.R., Fearnside, D., Curtis, T.P., et al. (2003) A comparative study of ammonia-oxidizing bacteria in lab-scale industrial wastewater treatment reactors. *Water Sci Technol* **48**: 17–24.
- Schmidt, I., Sliekers, O., Schmid, M., Bock, E., Fuerst, J., Kuenen, J.G., et al. (2003) New concepts of microbial treatment processes for the nitrogen removal in wastewater. *FEMS Microbiol Rev* **27**: 481–492.
- Schramm, A. (2003) In situ analysis of structure and activity of the nitrifying community in biofilms, aggregates, and sediments. *Geomicrobiol J* **20**: 313–334.
- Schramm, A., De Beer, D., Gieseke, A., and Amann, R. (2000) Microenvironments and distribution of nitrifying bacteria in a membrane-bound biofilm. *Environ Microbiol* **2**: 680–686.
- Schreiber, F., Wunderlin, P., Udert, K.M., and Wells, G.F. (2012) Nitric oxide and nitrous oxide turnover in natural and engineered microbial communities: biological pathways, chemical reactions and novel technologies. *Front Microbiol* **3**: 1–24.
- Sliekers, A.O., Derwort, N., Gomez, J.L.C., Strous, M., Kuenen, J.G., and Jetten, M.S.M. (2002) Completely autotrophic nitrogen removal over nitrite in one single reactor. *Water Res* **36**: 2475–2482.
- Sliekers, A.O., Third, K.A., Abma, W., Kuenen, J.G., and Jetten, M.S.M. (2003) CANON and Anammox in a gas-lift reactor. *FEMS Microbiol Lett* **218**: 339–344.
- van der Star, W.R.L., Miclea, A.I., van Dongen, U.G.J.M., Muyzer, G., Picioreanu, C., and van Loosdrecht, M.C.M. (2008) The membrane bioreactor: a novel tool to grow anammox bacteria as free cells. *Biotechnol Bioeng* **101**: 286–294.
- Stoecker, K., Dorninger, C., Daims, H., and Wagner, M. (2010) Double labeling of oligonucleotide probes for fluorescence in situ hybridization (DOPE-FISH) improves signal intensity and increases rRNA accessibility. *Appl Environ Microbiol* **76**: 922–926.
- Sundberg, C., Al-Soud, W.A., Larsson, M., Alm, E., Yekta, S.S., Svensson, B.H., et al. (2013) 454 pyrosequencing analyses of bacterial and archaeal richness in 21 full-scale biogas digesters. *FEMS Microbiol Ecol* **85**: 612–626.
- Terada, A., Lackner, S., Tsuneda, S., and Smets, B.F. (2007) Redox-stratification controlled biofilm (ReSCoBi) for completely autotrophic nitrogen removal: the effect of co-versus counter-diffusion on reactor performance. *Biotechnol Bioeng* **97**: 40–51.
- Terada, A., Lackner, S., Kristensen, K., and Smets, B.F. (2010) Inoculum effects on community composition and nitrification performance of autotrophic nitrifying biofilm reactors with counter-diffusion geometry. *Environ Microbiol* **12**: 2858–2872.
- Terada, A., Zhou, S., and Hosomi, M. (2011) Presence and detection of anaerobic ammonium-oxidizing (anammox) bacteria and appraisal of anammox process for high-strength nitrogenous wastewater treatment: a review. *Clean Techn Environ Policy* **13**: 759–781.
- Throbäck, I.N., Enwall, K., Jarvis, A., and Hallin, S. (2004) Reassessing PCR primers targeting *nirS*, *nirK* and *nosZ* genes for community surveys of denitrifying bacteria with DGGE. *FEMS Microbiol Ecol* **49**: 401–417.
- Tsushima, I., Kindaichi, T., and Okabe, S. (2007) Quantification of anaerobic ammonium-oxidizing bacteria in enrichment cultures by real-time PCR. *Water Res* **41**: 785–794.
- Vázquez-Padín, J., Mosquera-Corral, A., Luis Campos, J., Méndez, R., and Revsbech, N.P. (2010) Microbial community distribution and activity dynamics of granular biomass in a CANON reactor. *Water Res* **44**: 4359–4370.
- Wang, R., Terada, A., Lackner, S., Smets, B.F., Henze, M., Xia, S., et al. (2009) Nitrification performance and biofilm development of co- and counter-diffusion biofilm reactors: modeling and experimental comparison. *Water Res* **43**: 2699–2709.
- Wertz, S., Poly, F., Le Roux, X., and Degrange, V. (2008) Development and application of a PCR-denaturing gradient gel electrophoresis tool to study the diversity of *Nitrobacter*-like *nrxA* sequences in soil. *FEMS Microbiol Ecol* **63**: 261–271.
- Wüst, P.K., Horn, M.A., and Drake, H.L. (2011) *Clostridiaceae* and *Enterobacteriaceae* as active fermenters in earthworm gut content. *ISME J* **5**: 92–106.
- Yu, Y., Lee, C., Kim, J., and Hwang, S. (2005) Group-specific primer and probe sets to detect methanogenic communities using quantitative real-time polymerase chain reaction. *Biotechnol Bioeng* **89**: 670–679.
- Zhang, T.C., and Bishop, P.L. (1994) Evaluation of tortuosity factors and effective diffusivities in biofilms. *Water Res* **28**: 2279–2287.
- Zumft, W.G. (1997) Cell biology and molecular basis of denitrification. *Microbiol Mol Biol Rev* **61**: 533–616.

## Supporting information

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

**Table S1.** Probes used for the detection of target organisms by in-situ fluorescent hybridization.

**Table S2.** Index of related taxonomy for each functional guild.

**Fig. S1.** Reactor performance and microbial community abundances during reactor operation (qPCR performed with primers targeting functional genes).

**Fig. S2.** Typical O<sub>2</sub>-microprofiles during aeration periods within an aeration cycle.

**Fig. S3.** Rarefaction curves of denoised sequences and shared and unique OTUs between triplicate samples.

**Fig. S4.** Phylogenetic tree constructed with all the identified AOB, NOB and AnAOB sequences.